

Temporal compression mediated by short-term synaptic plasticity

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Time scales of cortical neuronal dynamics range from few milliseconds to hundreds of milliseconds. In contrast, behavior occurs on the time scale of seconds or longer. How can behavioral time then be neuronally represented in cortical networks? Here, using electrophysiology and modeling, we offer a hypothesis on how to bridge the gap between behavioral and cellular time scales. The core idea is to use a long time constant of decay of synaptic facilitation to translate slow behaviorally induced temporal correlations into a distribution of synaptic response amplitudes. These amplitudes can then be transferred to a sequence of action potentials in a population of neurons. These sequences provide temporal correlations on a millisecond time scale that are able to induce persistent synaptic changes. As a proof of concept, we provide simulations of a neuron that learns to discriminate temporal patterns on a time scale of seconds by synaptic learning rules with a millisecond memory buffer. We find that the conversion from synaptic amplitudes to millisecond correlations can be strongly facilitated by subthreshold oscillations both in terms of information transmission and success of learning.

hippocampus | temporal coding | oscillations | tempotron

Time scales of cortical neuronal dynamics are mostly in the range of few milliseconds to hundreds of milliseconds and are imposed on through cellular properties (1, 2), the kinetics of synaptic transmission (3), and complex connectivity patterns (4). These time scales are well adjusted to the time window for the induction of synaptic changes via spike-timing-dependent synaptic plasticity (5–7), which leads to the hypothesis that synaptic plasticity provides a means of learning to discriminate and recognize distinct cellular activity patterns. In contrast to cellular phenomena, the time scales of behavioral and cognitive phenomena, such as navigation in a maze or short-term memory, are in the order of seconds or longer. How to transfer the resulting slow behaviorally evoked temporal patterns to synaptic long-term changes and enable the formation of long-term memories is, however, largely unclear.

The present article proposes a new mechanism that uses short-term synaptic plasticity to encode temporal stimulus properties via variable amplitudes of synaptic currents. The memory time scales of short-term synaptic plasticity are in the range from hundreds of milliseconds to several seconds (8, 9) and thus provide a potential memory buffer on a behavioral time scale. It is described how the distribution of synaptic amplitudes resulting from short-term plasticity can be efficiently transferred into a temporal spike code that represents the slow input patterns. This mechanism thus constitutes a temporal compression from seconds to milliseconds. Using experimental and modeling investigations, we demonstrate how the combination of short-term synaptic plasticity and subthreshold membrane potential oscillations serves to generate an information-efficient temporal spike code (*cf.* ref. 10). For hippocampal CA3 pyramidal cells *in vitro*, we measure the temporal range of delays between excitatory postsynaptic currents (EPSCs) and action potentials (APs) varying the amplitude of a simulated synaptic input. Next, using a computational model, we reproduce the results of the *in vitro* approach and calculate how much information about the

input amplitude is conveyed via AP timing. Finally, we verify that the proposed mechanism is suitable for learning and decoding in downstream structures.

Results

Range of Delays Between Simulated EPSCs and APs. Synaptically induced currents determine the timing of a neuronal APs. In general, a large excitatory input current elicits a faster increase of the neuronal membrane potential than a small one. Thus, strong inputs give rise to shorter spike latencies than weak inputs and, consequently, the amplitudes of EPSCs are encoded in the timing of the APs. In this context, we first determined the temporal coding capabilities of APs triggered by varying EPSC input: For that, we measured the AP responses of a CA3 pyramidal cell induced by simulated EPSC sequences with increasing amplitudes (Fig. 1). These EPSC sequences are to represent the prominent short-term facilitation of single mossy fiber synapses evoking EPSCs with amplitudes from few tens of picoamperes to hundreds of picoamperes (for EPSC kinetics and amplitudes, see *Materials and Methods* and refs. 11–13). In addition to the simulated EPSCs, we imposed subthreshold oscillations with a frequency of 9 Hz, which is in the hippocampal theta band (4–12 Hz). Hippocampal theta oscillations are apparent in the extracellular field potential of freely behaving animals (14) and are also reflected through oscillations of the cellular membrane potential in anesthetized (15, 16) and behaving animals (17). Several repetitions of the stimulus train evoked a reliable pattern of postsynaptic APs (Fig. 1*A1*). The time delay between the onset of the EPSC and the postsynaptic AP ranged from 10.7 ± 0.3 ms (mean \pm SEM) for high EPSC amplitudes to 63 ± 2 ms for low EPSC amplitudes (Fig. 1*B*). For $n = 10$ cells, the average time delay between the onset of the EPSC and the postsynaptic AP ranged from 11 ± 2 ms to 63 ± 2 ms, providing a temporal coding range of 51 ± 2 ms (Fig. 1*C1*), which is approximately half of a theta period. The temporal coding range with subthreshold oscillations is significantly enhanced (t test: $P < 10^{-4}$) compared with experiments without oscillations [Fig. 1 in [supporting information \(SI\) Appendix](#)].

Determinants of Temporal Coding. Given subthreshold membrane potential oscillations, the temporal range of AP delays is a result of the interplay between oscillatory input and EPSC. In particular, the

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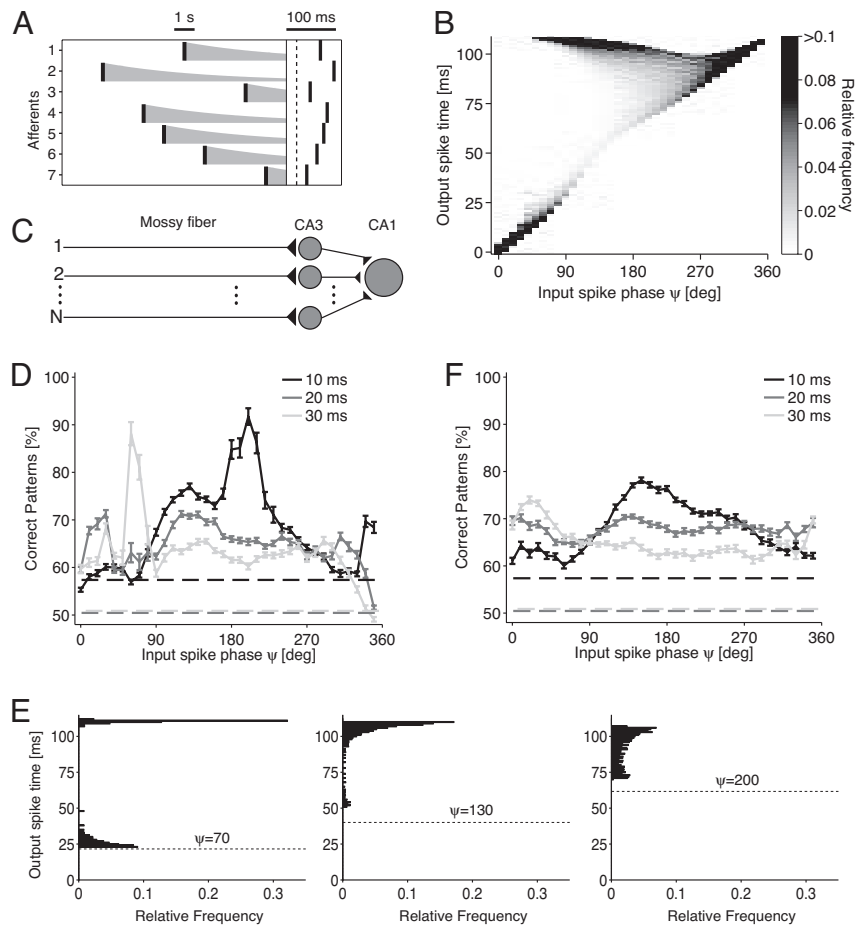


Fig. 3. Learning to discriminate between temporal patterns on a time scale of seconds. (A) Schematic illustration of temporal compression through synaptic facilitation. (Left) A temporal spike pattern (vertical bars) from a population of seven afferents on a time scale of seconds. The temporal pattern is preserved in the memory traces provided by the decay of synaptic facilitation (gray areas). (Right) Readout stimulus (dashed line) in combination with subthreshold oscillation evokes a reverse replay of the pattern in the seven neurons (vertical bars). Note different temporal scale bars for temporal pattern (Left) and reverse replay (Right). (B) Depending on the input phase ψ of the readout stimulus, the compressed population patterns show distinct phase distributions (gray-scale PSTHs). (C) A downstream threshold unit (e.g., in CA1) with adjustable synaptic weights learns to discriminate between temporally compressed phase patterns of a population of neurons (e.g., in CA3). (D) Fraction of correctly classified patterns after 100 learning cycles (mean \pm SEM, $n = 50$ simulations) reveals an optimal input phase ψ that depends on the decay time constant τ of the downstream EPSP (gray levels depict different values of τ). Dashed lines indicate fractions of correctly classified patterns for simulations without subthreshold oscillations. Standard errors for dashed lines are all smaller than 0.7%. (E) Exemplary PSTHs from B for input phases $\psi = 70^\circ$, 130° , and 200° (dashed lines), which yield best learning performance for the different time constants τ from D. (F) Adding a jitter (SD 30°) to the phase of the readout stimulus reduces performance of learning. Dashed lines are the same as in D, because there is no phase jitter without an oscillation.

tion is conveyed via the decaying slopes of the EPSPs. (ii) Learning can be moderately successful if the PSTH reveals a single peak that is narrower than the EPSP ($\psi = 130^\circ$, $\tau = 20$ ms). In this regime, the learning rule thus also extracts temporal information from the rising slope of the EPSP. (iii) Finally, successful learning occurs for bimodally peaked PSTHs ($\psi = 70^\circ$, $\tau = 30$ ms). In this case the temporal domain of the input pattern can be considered to be divided into two intervals (corresponding to high and low synaptic amplitudes) with peaked unimodal PSTHs each. As in the previous case, the temporal information with the two subgroups is again also conveyed via the rising slopes of the EPSPs.

In a final series of simulations, we jittered the phase of the readout stimulus (Fig. 3F). There, the general dependence on input phase is preserved compared with the noiseless case, although the peak performances are strongly reduced. Interestingly, the moderate performance peaks for a unimodal PSTH ($\psi = 130^\circ$) are almost unchanged.

To summarize, the success of the tempotron rule strongly depends on properties of the downstream neuron such as the decay

time constant τ of the EPSP. Moreover, using a model without subthreshold oscillations the success of learning is much smaller (dashed lines in Fig. 3D and F). Thus, subthreshold oscillations that increase the temporal coding range can improve or, as in the present example, may even be inevitable to enable learning.

Discussion

The present article elucidates how short-term synaptic plasticity can contribute to the temporal compression of slow behaviorally induced temporal patterns. The core idea is to map the temporal input pattern to a pattern of EPSC amplitudes. The memory time constant of synaptic short-term plasticity thereby determines the time window over which the slow temporal patterns can extend. To obtain a broad (high entropy) distribution of EPSC amplitudes, the interval distribution of presynaptic inputs has to match the time constant of short-term plasticity. Subthreshold oscillations with a period much shorter than the memory time scale of short-term plasticity have been used to efficiently translate the synaptic amplitudes into a temporal sequence of action potentials in a popu-

$$I = \int_{A_{\min}}^{A_{\max}} dA p(A) \int_0^{360^\circ} d\Phi p(\Phi|A) \lg_2 \left[\frac{p(\Phi|A)}{\int_{A_{\min}}^{A_{\max}} dA' p(\Phi|A') p(A')} \right]$$

between Φ and A . We therefore require a model of the amplitude distribution $p(A)$ and the conditional probability $p(\Phi|A)$ of having Φ , given A . The latter distribution is obtained through $p(\Phi|A) = \int_0^{360^\circ} d\psi p(\psi) \delta(\Phi - \Phi\psi(A))$, in which the phase transfer function $\Phi\psi(A)$ is derived from the two-compartment model (19) (see Fig. 2). The distribution $p(\psi)$ of input phases is assumed to be Gaussian with mean phase $\bar{\psi}$. Unless stated otherwise, the standard deviation of $p(\psi)$ was set to the exemplary value of 30° . The minimal and maximal EPSC amplitudes were $A_{\min} = 0 \frac{\mu A}{cm^2}$ and $A_{\max} = 14 \frac{\mu A}{cm^2}$, respectively.

Model. Computer simulations made use of a two compartment neuron model of a CA3 pyramidal cell by Pinsky and Rinzel (19). The model consists of a somatic and a dendritic compartment. The somatic compartment contains the standard Hodgkin–Huxley-type inactivating sodium and potassium delayed-rectifier currents. The dendritic compartment contains a calcium current, a potassium-mediated slowly activating afterhyperpolarization (AHP) current, and a fast activating potassium-mediated current, the saturation of which is calcium-dependent. The model has been extensively studied (e.g. ref. 38) and can be considered as one of the standard models for CA3 pyramidal cells.

The external currents applied to the model have the same shape as I_{osc} and I_{syn} in the electrophysiological recordings (see *Simulated EPSCs and Oscillations*). The amplitude of the oscillatory current was $I_1 = 2 \frac{\mu A}{cm^2}$, resulting in an amplitude of ≈ 5 mV for the membrane potential oscillation. The firing threshold was adjusted by an additional application of a constant current of $I_0 = -0.3 \frac{\mu A}{cm^2}$ to prevent firing for EPSP amplitudes $A = 0$. For simulations without subthreshold oscillations (dashed lines in Fig. 2D), we chose $I_1 = 0$ and $I_0 = -2.3 \frac{\mu A}{cm^2}$, which lead to a maximal delay of 70 ms between EPSC onset and AP. The model was implemented in C programming language. Integration was performed by using a fourth-order Runge–Kutta algorithm with adaptive step size (39).

- Netoff TI, et al. (2005) Synchronization in hybrid neuronal networks of the hippocampal formation. *J Neurophysiol* 93:1197–1208.
- Bendels MHK, Leibold C (2007) Generation of theta oscillations by weakly coupled neural oscillators in the presence of noise. *J Comput Neurosci* 22:173–189.
- Traub RD, Whittington MA, Colling SB, Buzsáki G, Jefferys JG (1996) Analysis of gamma rhythms in the rat hippocampus *in vitro* and *in vivo*. *J Physiol* 493:471–484.
- Memmesheimer RM, Timme M (2006) Designing the dynamics of spiking neural networks. *Phys Rev Lett* 97:188101.
- Markram H, Lübke J, Frotscher M, Sakmann B (1997) Regulation of synaptic efficacy by coincidence of postsynaptic APs and EPSPs. *Science* 275:213–215.
- Bi GQ, Poo M-M (1998) Synaptic modifications in cultured hippocampal neurons: Dependence on spike timing, synaptic strength, and postsynaptic cell type. *J Neurosci* 18:10464–10472.
- Kempster R, Gerstner W, van Hemmen JL (1999) Hebbian learning and spiking neurons. *Phys Rev E* 59:4498–4514.
- Zucker RS, Regehr WG (2002) Short-term synaptic plasticity. *Annu Rev Neurosci* 64:355–405.
- Nicoll R, Schmitz D (2005) Synaptic plasticity at hippocampal mossy fibre synapses. *Nat Rev Neurosci* 6:863–876.
- Hopfield JJ (1995) Pattern recognition computation using action potential timing for stimulus representation. *Nature* 376:33–36.
- Jonas P, Major G, Sakmann B (1993) Quantal components of unitary EPSCs at the mossy synapse on CA3 pyramidal cells of rat hippocampus. *J Physiol* 472:615–663.
- Toth K, Soares G, Lawrence JJ, Philips-Tansey E, McBain CJ (2000) Differential mechanisms of transmission at three types of mossy fiber synapse. *J Neurosci* 20:8279–8289.
- Gundfänger A, et al. (2007) Differential modulation of short-term synaptic dynamics by long-term potentiation at mouse hippocampal mossy fibre synapses. *J Physiol* 585:3:853–865.
- Buzsáki G (2002) Theta oscillations in the hippocampus. *Neuron* 33:325–340.
- Kamondi A, Acsády L, Wang XJ, Buzsáki G (1998) Theta oscillations in somata and dendrites of hippocampal pyramidal cells *in vivo*: activity-dependent phase-precession of action potentials. *Hippocampus* 8:244–261.
- Bland BH, Konopacki J, Dyck R (2005) Heterogeneity among hippocampal pyramidal neurons revealed by their relation to theta-band oscillation and synchrony. *Exp Neurol* 195:458–474.
- Lee AK, Manns ID, Sakmann B, Brecht M (2006) Whole-cell recordings in freely moving rats. *Neuron* 51:399–407.
- Thurley K, Leibold C, Gundfänger A, Schmitz D, Kempster R (2008) Phase precession through synaptic facilitation. *Neural Comput*, 10.1162/neco.2008.07.06-292.
- Pinsky PF, Rinzel J (1994) Intrinsic and network rhythmogenesis in a reduced Traub model for CA3 neurons. *J Comput Neurosci* 1:39–60.

Temporal Pattern Learning. We have defined 200 firing patterns of a population of 200 input lines such that each pattern contains one event (spike or burst) per input line occurring independently at random times t_i^n ($n = 1, \dots, 200$) in the interval between 0 and 10 seconds. Each input event triggers the maximal facilitation $A_{\max} = 14 \frac{\mu A}{cm^2}$ of the synapse connected to the specific input line. The facilitation f_n then decays exponentially with a time constant $\tau_f = 5$ seconds (Fig. 3A). To read out the states of facilitation, all synapses are simultaneously activated at a phase angle ψ with respect to an additional subthreshold oscillation with period 111 ms. The larger the delay between an input event and the readout stimulus, the smaller is the value of the facilitation f_n of the synapse at the time of readout. The readout stimulus therefore triggers a reverse replay of the pattern, and the replayed pattern is temporally compressed. The AP phases are obtained from the transfer function $\Phi\psi(A = f_n)$ depicted in Fig. 2C. The reversed and compressed pattern is then conveyed to a threshold unit and there elicits standardized EPSPs $w_n (e^{-\psi t} - e^{-\psi/\tau_s})$ (for $t \geq 0$) with a membrane time constant τ , a synaptic time constant $\tau_s = \tau/4$, and a synaptic weight w_n .

Half of the above patterns were randomly classified as “+” patterns, the others are termed “−” patterns. The synaptic weights are trained via the tempotron learning rule (23) such that the threshold unit emits a spike for + patterns and does not fire for − patterns. Unless stated otherwise, all parameters were taken as proposed by Gütiğ and Sompolinsky (23). In particular, in each learning cycle, all patterns were presented in the same order. Learning was stopped when 100% of the patterns were classified correctly or when the maximal number of learning cycles (here 100) was exceeded. Simulations were run for different EPSP time constants τ and different input phases ψ of the readout stimulus. We repeated simulations 50 times for each set of parameters. For the simulation results shown in Fig. 3D, the phase ψ of the readout stimulus was drawn from a normal distribution with mean $\bar{\psi}$ and standard deviation of 30° .

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- Mizumori SJ, McNaughton BL, Barnes CA (1989) A comparison of supramammillary and medial septal influences on hippocampal field potentials and single-unit activity. *J Neurophysiol* 61:15–31.
- Jung MW, McNaughton BL (1993) Spatial selectivity of unit activity in the hippocampal granular layer. *Hippocampus* 3:165–182.
- Salin PA, Scanziani M, Malenka RC, Nicoll RA (1996) Distinct short-term plasticity at two excitatory synapses in the hippocampus. *Proc Natl Acad Sci USA* 93:13304–13309.
- Gütiğ R, Sompolinsky H (2006) The tempotron: A neuron that learns spike timing-based decisions. *Nat Neurosci* 9:420–428.
- Hertz J, Krogh A, Palmer RG (1991) *Introduction to the Theory of Neural Computation*. (Westview, Boulder, CO).
- O’Keefe J, Recce ML (1993) Phase relationship between hippocampal place units and EEG theta rhythm. *Hippocampus* 3:317–330.
- Skaggs WE, McNaughton BL, Wilson MA, Barnes CA (1996) Theta phase precession in hippocampal neuronal populations and the compression of temporal sequences. *Hippocampus* 6:149–172.
- Mehta MR, Lee AK, Wilson MA (2002) Role of experience and oscillations in transforming a rate code into a temporal code. *Nature* 417:741–746.
- Harris KD, et al. (2002) Spike train dynamics predicts theta-related phase precession in hippocampal pyramidal cells. *Nature* 417:738–741.
- Huxter J, Burgess N, O’Keefe J (2003) Independent rate and temporal coding in hippocampal pyramidal cells. *Nature* 425:828–832.
- Gloveli T, Schmitz D, Empson R, Heinemann U (1997) Frequency-dependent information flow from the entorhinal cortex to the hippocampus. *J Neurophysiol* 78:3444–3449.
- Buzsáki G (1989) Two-stage model of memory trace formation: A role for “noisy” brain states. *Neuroscience* 31:551–570.
- Diba K, Buzsáki G (2007) Forward and reverse hippocampal place-cell sequences during ripples. *Nat Neurosci* 10:1241–1242.
- Foster DJ, Wilson MA (2006) Reverse replay of behavioural sequences in hippocampal place cells during the awake state. *Nature* 440:680–683.
- Buzsáki G, Draguhn A (2004) Neuronal oscillations in cortical networks. *Science* 304:1926–1929.
- Tsodyks MV, Markram HD (1997) The neural code between neocortical pyramidal neurons depends on neurotransmitter release probability. *Proc Natl Acad Sci USA* 94:719–723.
- Varela JA, et al. (1997) A quantitative description of short-term plasticity at excitatory synapses in layer 2/3 of rat primary visual cortex. *J Neurosci* 17:7926–7940.
- Dobrunz LE, Stevens CF (1999) Response of hippocampal synapses to natural stimulation patterns. *Neuron* 22:157–166.
- Booth V, Bose A (2001) Neural mechanisms for generating rate and temporal codes in model CA3 pyramidal cells. *J Neurophysiol* 85:2432–2445.
- Press WH, Teukolsky SA, Vetterling WT, Flannery BP (1992) *Numerical Recipes in C* (Cambridge Univ Press, Cambridge, UK).